Pilot study to assess the presence of Chlamydia trachomatis in urine from 18–30-year-old males using EIA/IF and PCR

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Abstract
Context. To increase detection, urine samples from young males could be opportunistically tested for Chlamydia trachomatis.
Objective. To determine C. trachomatis prevalence in urine, optimum specimen and compare sensitivity/feasibility of routine use of different testing methods.
Setting. Microbiology laboratory.
Samples. From males aged 18–30 years; group A = 71, group B = 83.
Main outcome measures. Chlamydia trachomatis positive if EIA- and IF- or PCR-positive.
Results. Group A: 12 EIA/IF-positive; 9/12 and 15 EIA-negative samples PCR-positive. Group B: 11 PCR-positive; 8/11 showed ‘sterile’ pyuria.
Conclusions. Opportunistic testing of urine from young men shows a significant number of C. trachomatis infections. ‘Sterile’ pyuria samples are optimal. EIA/IF are less sensitive than PCR but can be routinely performed and detect a significant proportion of cases.

Introduction
Chlamydia trachomatis infection can cause significant morbidity and any strategy that can increase detection rate should be exploited. Recent emphasis has been on screening women,1 however urine samples from young males could be readily utilised for opportunistic testing.2

We set out to assess C. trachomatis prevalence in urine samples sent routinely to our laboratory from males in the highest risk (18–30 years old) group, and to determine which type of specimen gave the greatest yield of positive results. It was assumed that medical consultation with submission of a urine specimen was a request to identify any significant pathogens. We aimed to compare our routine antigen detection methods [enzyme immunoassay/imunofluorescence (EIA/IF)] with polymerase chain reaction (PCR), which although regarded as having greater sensitivity requires batch testing and is significantly more expensive.

Methods
Between June 1998 and January 1999, 71 ‘sterile’ pyuria samples (>20 white cells/mm3 plus negative routine bacterial culture) from males aged 18–30 years (group A) were processed further by EIA for C. trachomatis antigen detection (MicroTrak®II). Sixty-two (87%) samples had clinical details of urinary tract symptoms or abnormal dipstick analysis. Fifty-one (72%) samples were received from general practitioners (GPs) and 17 (24%) from hospital inpatients. Samples weakly or clearly positive were further tested by IF (MicroTrak®). Samples were then stored at −20°C. Referring clinicians were telephoned about specimens that were EIA- and IF-positive. Ideal follow-up was discussed, i.e. endourethral samples for C. trachomatis, screening for other sexually transmitted diseases (STDs) and need for contact tracing; referral to the genitourinary medicine (GUM) clinic was advised. PCR was performed on the 71 stored undiluted samples using the ABI PRISM 7700 Sequence Detection System (PE Applied Biosystems, Warrington, UK). Samples that were PCR-negative but EIA+/−IF-positive were re-tested at a 1:10 dilution to reduce the effect of urine PCR inhibitors. PCR was also performed on 83 consecutive unslected urine samples from 18–30-year-old males collected between October 1998 and January 1999 (group B). Fifty-seven (69%) samples were received from GPs and 17 (21%) from hospital inpatients. Sixty-nine (83%) samples had clinical details of urinary tract symptoms and 24 (29%) showed ‘sterile’ pyuria.

Results
EIA and IF were positive for 12/71 (17%) patients. PCR detected 15/71 (21%) further positive urine samples, increasing the total yield to 27/71 (38%). However, three samples positive by EIA/IF were PCR-negative and five samples were only PCR-positive after being re-tested at a 1:10 dilution, consistent with the presence of PCR inhibitors. Eleven (13%) of the 83 group B samples were PCR-positive; eight of these (73%) demonstrated ‘sterile’ pyuria.

Discussion
Although recent emphasis has been on screening women, urine samples from males aged 18–30 years can be readily...
tested opportunistically for *C. trachomatis*. Although EIA/IF may have reduced sensitivity compared to PCR,³ our laboratory staff are experienced with these techniques and results are available on a same-day basis. Following this procedure, 17% of group A samples were found to be *C. trachomatis*-positive, three samples of which would have been missed by PCR alone. Furthermore, results are delayed with PCR as specimens are batch-tested for increased cost-effectiveness and inhibitors present in urine⁴ can lead to false-negative results unless diluted out. PCR detected *C. trachomatis* in 13% of group B urine samples (most showing ‘sterile’ pyuria) compared to 34% of group A urine samples, suggesting that resources are best deployed in testing ‘sterile’ pyuria samples. These results also suggest that *C. trachomatis* infection should be strongly considered in young men whose urine samples show ‘sterile’ pyuria. However, as 2/83 group B samples from symptomatic men without pyuria were PCR-positive, it is a further reminder that *C. trachomatis* should be considered in any young male complaining of dysuria.

Based on the present results we have introduced *C. trachomatis* EIA/IF testing of ‘sterile’ pyuria samples from young men aged 18–30 years as a routine laboratory investigation.

**Conclusions**

Opportunistic testing of ‘sterile’ pyuria samples from young men for *C. trachomatis* shows a significant yield of positive results that may otherwise have remained undetected. Although PCR is more sensitive than EIA/IF, the results may be delayed through batch testing, the procedure cannot be routinely performed in all laboratories, and it is significantly more expensive.

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**References**